

HYPERACCUMULATION OF NI BY *ALYSSUM MURALE* WALDST. & KIT. FROM ULTRAMAFICS IN BOSNIA AND HERZEGOVINA

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Abstract. We investigate the nickel concentration in serpentine soil samples from several areas in central region of Bosnia and Herzegovina. Concentration of some other elements such as P, K, Ca, Mg, Fe, Mn, Pb, Co, and Cd were determined as well. Selected soil samples were typical of ultramafic sites with low concentrations of P, K and Ca and high concentrations of Mg, Fe, Ni and Zn. The Mg/Ca ratio was relatively high (0.78–30.61). The levels of P, K, Fe and Zn were high, Mn and Cu occurred in low amounts, whereas, Cd, Co and Pb were only traceable. Principal component analysis pointed out the correlation of elements originating from ultramafic bedrock and their possible phytoextraction. The concentration of Ni in tissues of all investigated plant samples was high to extremely high. High concentrations of Co in plant tissues were recorded also. Bearing in mind that *A. murale* at investigated sites in Bosnia and Herzegovina showed a high capacity for accumulation and translocation of Ni and generally, shared high tolerance to other metals, this plant may be used to clean up soils contaminated with heavy metals, as well as for phytomining.

Keywords: *serpentine soil, nickel, soil contamination, heavy metal cleanup, phytomining, phytoextraction*

Introduction

Ultramafics are defined as group of magmatic or metamorphic rocks that consist of less than 45 % silica (SiO₂) and have high concentrations of magnesium (Mg), iron (Fe), chrome (Cr), cobalt (Co) and nickel (Ni). In addition to high concentrations of those elements, ultramafic soils, developed by weathering of ultramafic bedrock, are generally characterized with low concentrations of phosphorus (P), potassium (K), and calcium (Ca) (Alexander et al., 2007). During the serpentine creation, the main process is metamorphism (also known as metasomatism) of primary magnesium-iron silicate minerals (Alexander et al., 2007). In the course of this process primary silicate minerals are replaced by magnesium silicate serpentine minerals (Alexander et al., 2007). Ultramafics exist world-wide, and in Europe they are concentrated on the area of Balkan Peninsula (Stevanović et al., 2003). Weathering of serpentinite and serpentine rocks differ from site to site as a result of different climatic conditions in addition to the nature of parent material and other factors such as topography, biota and time (Oze et al., 2004).

Serpentines are very special plant-soil-rocks system which are characterized with the growth of specific and extraordinary plant species, and support vegetation in remarkable physiognomic contrast with that on other soil areas (Whittaker, 1954; Lazarus et al.,

2011). Some of these characteristics are related to the fact that ultramafic soil is largely sterile and unproductive, and if the plants flourish in such harsh environment, they are generally endemic species. The infertility of these soils is characterized as serpentine unproductiveness (Karataglis et al., 1982; Jackson, 2005). A large number of plants that flourish on serpentine soil belongs to the species that are referred to as “hyperaccumulators” (Van der Ent et al., 2013). These plants can take up metal and metalloid trace elements and store them in their leaves, flowers and stems in remarkably high concentrations, thus providing an adaptation for growth on soils that otherwise might be toxic. More than 500 species of “hyperaccumulator” have been indentified, including a significant number of representatives of the family Brassicaceae (Pollard et al., 2014).

According to Broadhurst and Chaney (2016), within the family Brassicaceae, the genus *Alyssum* contains the largest number of ascertained Ni hyperaccumulators – 50 taxa, including the species *Alyssum murale* Waldst. & Kit. which is distributed in southeastern Europe, Russia, Asia Minor and southwestern Asia (Ball and Dudley, 1993). In relation to the recent molecular, genetic and morphological evidence, from native populations, several species previously regarded as endemic hyperaccumulators can hardly be accepted as separate from *A. murale*, and we should treated all these taxa as synonyms of the species *A. murale* (Reeves et al., 2001; Hartvig, 2002; Whiting et al., 2003). *A. murale* is widespread in the serpentines in the Balkans and its accumulation potential is well documented in Serbia (Tumi et al., 2012) and, in particular, in Albania, Greece and Bulgaria (Bani et al., 2007; 2009; 2010; 2013). Moreover, *A. murale* is reported to have a remarkable potential for application in remediation of metals in the environment, as well as a for nickel-phytomining (Li et al., 2003; Bani et al., 2015; Nkrumah et al., 2016).

Given the importance of *A. murale* as a potential source for a phytomining and the fact that there are no data on hyperaccumulation characteristic of its native populations in Bosnia and Herzegovina, nor on the soil characteristics over a wide serpentinite area in this region, the aims of this study were: (a) to investigate soil chemical properties from several serpentine sites in central region of Bosnia and Herzegovina on which this species was recorded; (b) to investigate levels of accumulation and translocation of trace metals in plant tissues (roots, stems, and leaves), with a specific focus on Ni; and, (c) to estimate the potential use of local population for phytoextraction.

Materials and Methods

Studied area

Studied area covered dispersed serpentine outcrops in central part of Bosnia and Herzegovina, covering the surface of cca. 200 km². Samples were taken from 10 different locations, i.e. sampling points (SP1-SP10) based of presence of large populations of *A. murale* at each site (Table 1, Fig. 1). These locations were mainly in the uninhabited locations. The low amount of topsoil is mainly derived from weathered serpentinite. The serpentine soil on all sites are rocky and very shallow, vulnerable to erosion They are characterised mainly of sparse vegetation in the vicinity of pine and oak forests.

Sample collection

A. murale and soil samples were collected from May 2013 to May 2014 from 10 sampling points (SP), SP1-SP10 (Table 1, Fig. 1). Soil samples (cca. 500 g per sample) were taken from the main rooting area on each site, under all field-collected specimens of *A. murale*. Soil samples were first air-dried at room temperature, and thereafter pulverized, sieved through the 50 µm sieve and oven-dried overnight.

Plant samples (cca. 500 g per sample) were collected from each of the investigated location, separated into roots, stems and leaves and prepared for the analysis according to the procedure described by Markert (1995). Voucher specimens of all the plants material collected are deposited in the Collections of the Institute of Botany and Botanical Garden, Faculty of Biology, University of Belgrade (BEOU).

Table 1. Locations of the sampling points of *Alyssum murale* populations from Bosnia and Herzegovina

Location	Sample point	Latitude (N)	Longitude (E)
Bakotic	SP 1	44°32'22.3"	18°10'27.9"
BosanskoPetrovo	SP 2	44°37'47.0"	18°20'31.2"
Causevica	SP 3	44°29'19.8"	18°10'57.8"
Klokotnica	SP 4	44°43'17.8"	18°09'51.7"
Liplje	SP5	44°38'11.9"	17°31'52.5"
Maglaj	SP 6	44°32'21.6"	18°05'19.6"
Stanari	SP 7	44°42'19.3"	17°46'47.7"
Sehar	SP 8	44°26'25.8"	18°06'51.9"
Teslić	SP 9	44°36'58.1"	17°49'29.1"
Zvornik	SP 10	44°24'00.1"	19°06'38.1"

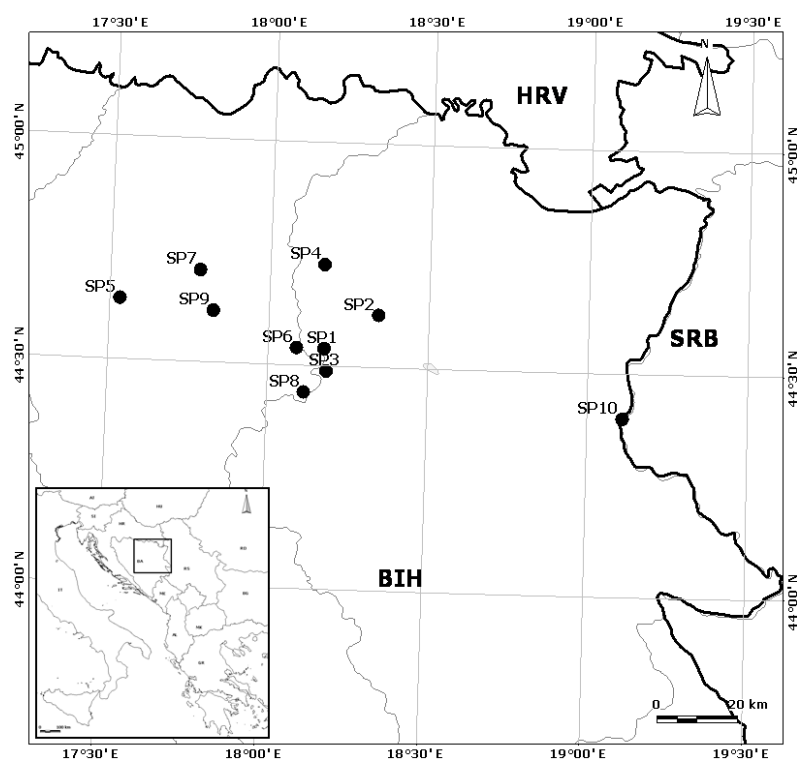


Figure 1. Map of the areas in Bosnia and Herzegovina where soil and plant samples were collected

Soil analysis

The distribution of the particle sizes was determined after soil dispersion by sodium hexametaphosphate, using the pipette method for the silt and clay fractions and dry-sieving for the sand fraction (Van Reeuwijk, 1995). The sand particles are separated from clay and silt with a 50 µm filter. Determination of soil textural classes was done following the USDA classification (Rowell, 1997). pH of soil samples (actual and exchangeable) was determined in distilled water and in 1 M KCl solution, respectively, in a solid-liquid (S/L) ratio of 1:2.5 ml/g (McKeague, 1978). Concentration of available P₂O₅ and K₂O were measured in AL solution (0.1 M ammonium lactate and 0.4 M acetic acid) extract (S/L 1:20) (Egner et al., 1960). Content of phosphate was determined by molybdenum blue method and potassium concentration was determined using FES (flame emission spectrophotometry) by Pye Unicam SP 192 atomic absorption spectrophotometer. Concentration of available Ca and Mg were determined in 1 M ammonium acetate extract (S/L 1:50) using AAS (atomic absorption spectrophotometry) (Pye Unicam SP 192) (Van Reeuwijk, 2002). Dichromate digestion based on FAO procedure was used for determination of organic matter concentration (FAO, 1974). Concentration of available (potentially leachable) metals in soil samples was determined by 0.1 N HCl (S/L 1:10) according to the procedure recommended by Garcia et al. (1979). Total metal extraction was done by HCl and HNO₃ digestion (ISO 11466 1995). Metal concentrations in investigation extracts were determined using atomic absorption spectrophotometry (ISO 11047 1998) (Pye Unicam SP 192). Each analysis of the soil material was performed in triplicates as well as each chemical analysis.

Plant analysis

Dried and powdered plant material from *A. murale* was digested by slightly modified wet procedure described by ISO 6636/2, 1981, using a boiling mixture of nitric and sulphuric acids. Concentration of Ca in plant tissue samples was determined using FES (flame emission spectrophotometry) using a Pye Unicam SP 192 atomic absorption spectrophotometer. Metal concentrations were determined using atomic absorption spectrophotometry (Pye Unicam SP 192). The series of standard solutions for metals were made from 1 g/l solutions purchased from Carlo Erba, Italy. Each analysis of the plant samples was performed in triplicates. In addition, each of the chemical analyses was also run with three replicates.

Data analysis

The results of soil and plant analyses are expressed as mean ± standard deviation (SD). Normality was checked for all measured variables (Shapiro-Wilks test, $\alpha=0.05$), whereupon they were log transformed prior to further analysis. A principal components analysis (PCA) based on correlation matrix was done in order to reveal the relationship between analysed serpentine sites concerning the element concentrations. Measured variables used in PCA were mean-centred and standardized. The number of principal components (PCs) that were extracted from the variables was defined by Kaiser's criterion, ie. only PCs with eigenvalues higher than 1 were retained (Kaiser and Rice, 1974). Values of element concentrations in plant tissues, as well as between concentration of available elements in the soil samples and those in plants were

compared by Pearson's correlation test using the program SPSS 19 for Windows. Differences at $p < 0.05$ were accepted as the level of significance.

For the evaluation of trace metal tolerance of *A. murale*, two indices were determined, bioconcentration factor (BCF) and translocation factor (TF) (Ghosh and Singh, 2005). Bioconcentration factor which indicates the overall ability of plant to accumulate trace metals was calculated as the quotient of their concentrations in shoots and the available (potentially leachable) concentrations in the soil. The translocation factor, indicating the plant ability to translocate trace metals from the root to the aerial parts, was calculated as the ratio of their concentrations in shoots and roots.

Results

Soil characteristics

The analysed chemical properties of 10 investigated soil samples, such as pH, organic matter, P_2O_5 , K_2O , available concentrations of major elements (Fe, Ca, Mg), and total and available concentrations of trace elements (Ni, Mn, Zn, Cu, Cr, Co, Cd, Pb), are outlined in *Table 2*. The pH of the soil samples diverse from moderately acidic to moderately alkaline, while pH in 1 M KCl was acidic to almost neutral. The percentage of organic matter differed within a moderate range. In regard to the concentrations of analysed elements, soil samples were more or less typical of the soil that is developed on the ultramafic rocks. Soils at the investigated sites had moderate to high concentrations of Mg, and low to moderate concentrations of Ca, thus having Mg/Ca quotient >1 in all but one location (SP3). Furthermore, all examined soil samples had a high concentration of trace elements that are typical for serpentinite locations (Ni, Co, Cr, Mn), while the rest of the analysed elements were in the ranges of normal soils.

Chemical composition of the plant material

Concentrations of analysed elements in the roots, stems, and leaves of 10 analysed *A. murale* populations from sampling locations are presented in *Table 3*. Regarding trace metals which are typical for serpentinites, the highest concentrations of Ni and Mn were recorded in leaves, and the lowest in roots of analysed plants. The same trend was also noticed for Co, which is recorded in the extremely high concentrations (up to $142.9 \text{ mg} \cdot \text{kg}^{-1}$ in leaves of *A. murale*). The opposite trend was noticed for Zn, since the concentration of Zn was the lowest in the leaves, while in both, in the stems and in the roots, it was significantly higher. Concentration of Cd was very low and similar in all investigated samples of *A. murale*. Only in several leaf samples Cd concentration was slightly higher. Pb was not detected in plant in one location, and in all others Pb concentration was relatively low, especially in roots.

Table 2. pH, organic matter(%), P_2O_5 , K_2O , major elements (Fe, Ca, Mg) and trace elements (Ni, Mn, Zn, Cu, Cr, Co, Cd, Pb) concentrations (mg kg⁻¹) in soils of investigated location in which *Alyssum murale* are present (mean ±SD)

Sample point	SP1	SP2	SP3	SP4	SP5	SP6	SP7	SP8	SP9	SP10
pH soil in H ₂ O	8.2	8.1	6.57	7.91	7.56	8.6	6.91	8.29	8.37	8.05
pH soil in 1 N KCL	7.37	7.33	5.56	6.92	6.16	7.49	6.91	6.96	7.21	6.9
Organic matter %	0.82±0.09	1.99±0.15	4.85±1	1.89±0.1	1.98±0.1	2.2±0.06	1.05±0.1	0.5±0.1	1.24±0.07	1.3±0.1
P ₂ O ₅ (a)	1.72±0.2	30.8±0.3	3.85±0.6	2.47±0.5	1.37±0.2	10.5±0.5	5.06±0.7	1.66±0.5	2.87±0.4	2.58±0.4
K ₂ O (a)	9.42±0.8	4.18±0.3	50.3±1.9	9.9±0.06	9.17±0.1	20.38±1.1	35.42±1.3	5.2±0.03	13.41±0.6	11.8±0.6
Fe (a)	777.7±19	780.4±7.1	502.1±6.6	142.1±1.3	678.6±11	849.5±11.6	387.7±3.3	248.4±12.5	331.5±3.6	507.5±20.7
Ca (a)	1055.6±29	222.9±16	527.2±36	317.9±8.8	186.8±6.7	2419.6±120	414.6±16	112.1±8.6	687.1±51	1846.6±32
Mg (a)	2422.3±56	656.1±41	411.2±32	925.7±9.7	883.1±9.8	4257.6±76	3631±100	3429.7±330	2198±135	3312.2±123
Mg/Ca	2.29	2.94	0.78	2.91	4.73	1.76	8.76	30.61	3.20	1.79
Ni (t)	2232.1±50	1675.2±38	637.2±9.3	2075.4±12	1077.4±38	2222.5±99	1555.6±28	1862.2±29	2052.3±16	2222.3±42
Ni (a)	2007.5±4.4	1131.9±2.0	325.4±7.5	1884.7±2.9	1633.9±68	1456.4±24	1296.9±1.4	1873.1±0.7	1158.3±5.1	2036.2±41
Mn (t)	1577.6±10	1772.6±102	1426.1±69	1735.4±54	1875.7±47	1458.8±15.0	1298.5±6.1	1884.3±105	2540.1±10.7	2442.7±65
Mn (a)	590.5±6.55	548.8±4.5	508.9±1.88	426.6±15.1	648.3±1.40	479.8±6.39	476.3±9.23	150.8±9.61	712.6±0.93	860.0±21.6
Zn (t)	107.78±4.7	36.96±3.3	101.1±1.8	48.53±0.89	88.08±0.9	36.8±2.4	106.3±1.01	95.9±7.5	120.47±5.2	56.49±0.61
Zn (a)	26.81±2.79	1.72±0.21	12.01±0.58	1.70±0.18	11.51±0.44	1.53±0.16	13.67±2.14	36.35±2.52	15.13±2.2	2.36±0.34
Cu (t)	0.809±0.01	2.35±0.32	7.52±0.16	5.17±0.01	27.15±1.10	2.52±0.27	28.51±1.46	3.55±0.14	21.49±0.84	9.39±0.14
Cu (a)	0.97±0.04	0.516±0.04	<UDL	<UDL	<UDL	<UDL	1.47±0.02	<UDL	<UDL	3.33±0.04
Cr (t)	153.9±26	1748.2±21	837.4±15	674.5±15	539.3±11	267.5±36	798.7±11	402.6±10	305.7±1.09	804.2±8.0
Cr (a)	0.53±0.2	47.31±0.03	<UDL	15.79±1	5.14±0.33	4.13±0.33	17.44±0.16	9.34±0.03	<UDL	<UDL
Co (t)	243.1±9.8	201.4±6.4	76.38±12	211.8±12	224.3±17	229.2±15	234.4±11	296.8±18	200.2±14.9	204.8±14
Co (a)	9.31±2.2	21.21±1.5	20.56±1.9	73.07±10.5	65.37±4.3	49.67±4.5	55.19±6.9	73.66±2.9	51.48±3.9	86.92±6.5
Cd (t)	1.34±0.14	3.38±0.21	3.83±0.05	<UDL	3.58±0.06	<UDL	1.88±0.008	<UDL	2.29±0.26	2.88±0.02
Cd (ae)	0.52±0.001	1.47±0.12	<UDL	<UDL	<UDL	<UDL	0.435±0.12	<UDL	0.262±0.001	1.34±0.02
Pb (t)	17.59±0.02	46.93±0.97	128.09±0.47	86.97±0.42	11.41±0.02	13.68±0.05	71.52±0.15	32.06±0.44	21.07±0.97	128.7±0.02
Pb (a)	<UDL	<UDL	12.70±0.01	0.679±0.05	<UDL	<UDL	1.42±0.03	<UDL	4.71±0.01	37.21±0.20

Abbreviation used: (t) – total; (a) – available; <UDL – under the detection limit

Table 3. Concentrations of analysed elements in *Alyssum murale* roots, stems and leaves (mean $\pm$ SD)

Sample point	SP1	SP2	SP3	SP4	SP5	SP6	SP7	SP8	SP9	SP10
Fe root	319.1 $\pm$ 12	885.2 $\pm$ 8.6	2328.7 $\pm$ 251	480.2 $\pm$ 1.2	435.8 $\pm$ 25	471.6 $\pm$ 14	122.2 $\pm$ 1.9	364.9 $\pm$ 5.7	461.3 $\pm$ 5.7	547.7 $\pm$ 179
Fe stem	144.7 $\pm$ 14	1159.1 $\pm$ 41	1600.8 $\pm$ 157	386.7 $\pm$ 4.9	763.5 $\pm$ 43	279.3 $\pm$ 4.1	306.3 $\pm$ 5.0	685.2 $\pm$ 7.0	835.8 $\pm$ 35	637.1 $\pm$ 21
Fe leaves	213.1 $\pm$ 22	1454.1 $\pm$ 39	1172.8 $\pm$ 47	755.9 $\pm$ 3.7	928.4 $\pm$ 12	637.3 $\pm$ 39	334.7 $\pm$ 13	964.7 $\pm$ 35	562.4 $\pm$ 26	444.5 $\pm$ 17
Ca root	8273.3 $\pm$ 64	5323.5 $\pm$ 3.7	3243.5 $\pm$ 40	7343.9 $\pm$ 26	8385.5 $\pm$ 42	5138.6 $\pm$ 99	3819.5 $\pm$ 23	4417.3 $\pm$ 31	7757.1 $\pm$ 26	8653.1 $\pm$ 20
Ca stem	41419.2 $\pm$ 42	29255.8 $\pm$ 18	19170.1 $\pm$ 17	22276.4 $\pm$ 11	11362.6 $\pm$ 9.5	33454.6 $\pm$ 18	9849.2 $\pm$ 19	7233.6 $\pm$ 31	17569.9 $\pm$ 6.6	19504.5 $\pm$ 3.8
Ca leaves	61169.2 $\pm$ 1.9	42197.7 $\pm$ 3.9	41132.0 $\pm$ 10	55115.8 $\pm$ 4.2	17283.7 $\pm$ 16	53660.3 $\pm$ 25	22733.3 $\pm$ 40	13104.2 $\pm$ 24	42344.2 $\pm$ 33	33458.6 $\pm$ 36
Mg root	2184.1 $\pm$ 41	5283.5 $\pm$ 34	2537.4 $\pm$ 365	2522.6 $\pm$ 4.7	2367.6 $\pm$ 245	1158.7 $\pm$ 12	2052.2 $\pm$ 169	5157.6 $\pm$ 395	956.9 $\pm$ 7.2	1669.3 $\pm$ 232
Mg stem	2608.6 $\pm$ 217	5363.2 $\pm$ 33	3766.7 $\pm$ 31	3111.4 $\pm$ 6.3	3076.8 $\pm$ 321	2866.2 $\pm$ 453	2840.5 $\pm$ 326	8250.5 $\pm$ 400	1715.3 $\pm$ 66	2855.9 $\pm$ 337
Mg leaves	9686.3 $\pm$ 150	8200.7 $\pm$ 14	9620.3 $\pm$ 110	5220.9 $\pm$ 6.3	5337.4 $\pm$ 538	6251.9 $\pm$ 647	6457.4 $\pm$ 528	10103.4 $\pm$ 31	2974.1 $\pm$ 323	4541.1 $\pm$ 411
Ni root	1009.9 $\pm$ 6.1	1808.9 $\pm$ 14	1738.6 $\pm$ 47	983.9 $\pm$ 3.1	2386.7 $\pm$ 52	1123.8 $\pm$ 48	2504.7 $\pm$ 356	2076.2 $\pm$ 178	2647.7 $\pm$ 67	5885.4 $\pm$ 11
Ni stem	3314.3 $\pm$ 116	2400.9 $\pm$ 2.1	2885.7 $\pm$ 349	1336.8 $\pm$ 45	2838.1 $\pm$ 178	2323.8 $\pm$ 13	2938.1 $\pm$ 24	4219.1 $\pm$ 178	3742.8 $\pm$ 233	7409.5 $\pm$ 178
Ni leaves	9695.2 $\pm$ 178	7821.3 $\pm$ 1.5	4457.1 $\pm$ 308	2304.5 $\pm$ 2.4	6028.6 $\pm$ 308	7457.1 $\pm$ 617	6504.7 $\pm$ 293	8933.3 $\pm$ 793	5481 $\pm$ 299	10552.4 $\pm$ 242
Mn root	38.16 $\pm$ 3.3	22.38 $\pm$ 0.6	30.76 $\pm$ 2.4	11.88 $\pm$ 0.4	24.29 $\pm$ 1.3	100.5 $\pm$ 2.5	14.88 $\pm$ 2.8	7.06 $\pm$ 2	18.92 $\pm$ 0.8	44.59 $\pm$ 2.2
Mn stem	17.91 $\pm$ 1.9	33.19 $\pm$ 1.6	15.18 $\pm$ 2.5	63.51 $\pm$ 1.2	39.11 $\pm$ 4.9	99.03 $\pm$ 0.7	36.21 $\pm$ 4.3	16.73 $\pm$ 3.3	19.86 $\pm$ 0.9	66.99 $\pm$ 1.9
Mn leaves	97.61 $\pm$ 7.8	43.52 $\pm$ 0.4	109.6 $\pm$ 6.8	71.85 $\pm$ 0.4	95.01 $\pm$ 2.4	135.0 $\pm$ 0.5	101.1 $\pm$ 7.8	36.25 $\pm$ 2	71.26 $\pm$ 5.3	107.5 $\pm$ 5.9
Zn root	26.09 $\pm$ 2.2	8.28 $\pm$ 0.0002	2.62 $\pm$ 0.1	8.09 $\pm$ 0.1	2.46 $\pm$ 0.1	1.72 $\pm$ 0.2	5.64 $\pm$ 0.3	7.44 $\pm$ 0.7	7.26 $\pm$ 0.2	9.81 $\pm$ 0.4
Zn stem	13.21 $\pm$ 0.5	9.86 $\pm$ 0.7	3.24 $\pm$ 0.1	2.14 $\pm$ 0.7	5.37 $\pm$ 1.9	4.45 $\pm$ 0.2	1.46 $\pm$ 0.1	1.69 $\pm$ 0.3	0.61 $\pm$ 0.2	2.64 $\pm$ 0.04
Zn leaves	4.34 $\pm$ 0.2	11.28 $\pm$ 0.01	3.04 $\pm$ 0.6	2.62 $\pm$ 0.4	4.33 $\pm$ 0.5	0.51 $\pm$ 0.1	0.49 $\pm$ 0.09	0.35 $\pm$ 0.09	1.17 $\pm$ 0.06	1.25 $\pm$ 0.3
Co root	30.61 $\pm$ 8.3	41.20 $\pm$ 0.82	11.96 $\pm$ 1.32	13.16 $\pm$ 0.16	23.94 $\pm$ 12	12.10 $\pm$ 0.80	115.5 $\pm$ 5.0	56.49 $\pm$ 4.0	54.28 $\pm$ 4.9	55.81 $\pm$ 4.1
Co stem	73.05 $\pm$ 8.6	52.89 $\pm$ 0.50	19.63 $\pm$ 0.72	20.91 $\pm$ 1.21	11.89 $\pm$ 1.31	58.29 $\pm$ 5.3	19.51 $\pm$ 0.6	11.93 $\pm$ 1.3	12.03 $\pm$ 1.4	38.87 $\pm$ 1.4
Co leaves	142.8 $\pm$ 8.3	81.36 $\pm$ 0.64	122.2 $\pm$ 8.3	38.05 $\pm$ 0.08	20.61 $\pm$ 0.16	39.84 $\pm$ 0.76	51.76 $\pm$ 1.05	115.6 $\pm$ 17	41.22 $\pm$ 1.1	19.29 $\pm$ 0.8
Cd root	1.87 $\pm$ 0.25	1.25 $\pm$ 0.04	0.652 $\pm$ 0.4	2.72 $\pm$ 0.06	1.39 $\pm$ 0.51	1.03 $\pm$ 0.53	2.63 $\pm$ 1.0	0.896 $\pm$ 0.27	2.06 $\pm$ 0.41	1.41 $\pm$ 0.37
Cd stem	1.76 $\pm$ 0.9	1.8 $\pm$ 0.02	2.41 $\pm$ 0.18	3.21 $\pm$ 0.08	2.81 $\pm$ 0.17	3.63 $\pm$ 0.36	2.20 $\pm$ 0.21	2.66 $\pm$ 0.38	1.709 $\pm$ 0.64	2.14 $\pm$ 0.16
Cd leaves	3.47 $\pm$ 0.6	2.42 $\pm$ 0.25	2.22 $\pm$ 0.23	4.30 $\pm$ 0.28	2.38 $\pm$ 0.58	2.39 $\pm$ 0.40	2.00 $\pm$ 0.49	3.11 $\pm$ 0.57	0.582 $\pm$ 0.45	1.89 $\pm$ 0.29
Pb root	8.23 $\pm$ 0.4	2.21 $\pm$ 0.05	6.87 $\pm$ 0.4	<UDL	2.48 $\pm$ 0.2	3.82 $\pm$ 0.5	13.02 $\pm$ 0.6	16.59 $\pm$ 0.3	27.26 $\pm$ 0.4	14.17 $\pm$ 0.9
Pb stem	10.96 $\pm$ 0.7	1.34 $\pm$ 0.04	18.07 $\pm$ 0.6	<UDL	17.97 $\pm$ 0.7	8.00 $\pm$ 0.1	43.09 $\pm$ 3.8	26.78 $\pm$ 0.8	17.96 $\pm$ 0.07	5.60 $\pm$ 0.4
Pb leaves	17.30 $\pm$ 0.4	3.45 $\pm$ 0.3	16.59 $\pm$ 0.9	<UDL	8.57 $\pm$ 0.4	6.24 $\pm$ 0.4	27.16 $\pm$ 0.4	16.48 $\pm$ 1	17.83 $\pm$ 0.7	9.50 $\pm$ 0.3

Abbreviation used: <UDL – under the detection limit

Relationship between metal concentrations in soil and plants

The results of principal components analysis of the measured element concentrations in investigated soil samples are presented in *Table 4* and *Figure 2*. PCA rendered five principal components with eigenvalues greater than 1.00, accounting for 86.8% of the total variability of analysed data set. Taking into account these five factors, percentage of variable communalities ranged from 65.9% for Cu, up to 95.2% for K, so that they provide a good overview of all analyzed elements. The first component (PC1) explaining 30.0% of the total variance, showed a strong negative correlation with Ni, Co and Mg, and a moderate positive one with Cr and Cd. Namely, PC1 that was mostly determined by elements originating from parent rock, separated sites characterized by the highest concentrations of Ni, Co and Mg, with somewhat lower concentrations of Cr and Cd. The second component (PC2), which explained 18.8% of the total variance, showed strong positive correlation with P and strong negative one with Zn. Finally, the third, fourth and fifth principal components, explaining 15.9%, 11.8% and 10.2% of the total variance, were mostly determined by Ca, Fe and Mn concentrations, respectively.

Results of correlation analyses of metal concentrations in *A. murale* showed the strongest synergism for K – Pb in both roots and shoots, and the strongest antagonism for Fe – Zn in roots, and Ca – K in shoots (*Table 5*). Regarding the concentrations of metals in plant tissues (roots, stems and leaves) in relation to their available concentrations in soil samples, the strongest positive correlation was noticed between Ca concentration in soil and those in both leaves and stems, while the strongest negative one between Zn in soil and those in roots and stems (*Table 6*).

More detailed relations between available trace metals in soil and plant tissues can be observed from the calculated values of bioconcentration and translocation factor (*Table 7*). The obtained values of bioconcentration factors indicated a good uptake efficiency for all elements listed, except Mn for which the BCF was less than 1 in all investigated localities. In respect of translocation factor, efficient translocation is also observed, except for Zn which was mostly stored in root.

Table 4. Factor loadings, percentage of variable communalities, eigenvalue and percentages of variance for the first five principal components (PC1-PC5, factor loadings > 0.75 are indicated in bold)

Parameter	PC1	PC2	PC3	PC4	PC5	% of community
Ca	-0.2086	0.1846	0.7452	-0.1565	0.5311	94.0
Mg	-0.7528	-0.1875	0.2838	-0.1821	0.2931	80.1
P	0.1611	0.8735	0.0257	0.0552	0.0576	79.6
K	0.5334	-0.2465	0.6526	-0.4145	0.0941	95.2
Fe	0.0567	0.4404	0.6049	0.6220	0.0011	95.0
Ni	-0.8886	0.1626	-0.095	-0.1218	0.3285	94.8
Zn	0.1619	-0.8697	0.1610	0.1549	-0.1451	85.4
Mn	-0.2517	-0.2351	-0.4363	0.3687	0.6212	83.1
Cu	0.3906	-0.5875	-0.1119	0.1268	0.3641	65.9
Cr	0.6547	0.4068	-0.5171	-0.0721	0.225	91.7
Pb	0.5577	0.0880	-0.2478	-0.5973	0.3717	87.5
Cd	0.6818	-0.0455	0.0798	0.5932	0.3365	93.8
Co	-0.8609	-0.0399	-0.2653	0.1243	-0.0202	82.9
Eigenvalue	3.9	2.4	2.1	1.5	1.3	
% Total variance	30.0	18.8	15.9	11.8	10.2	

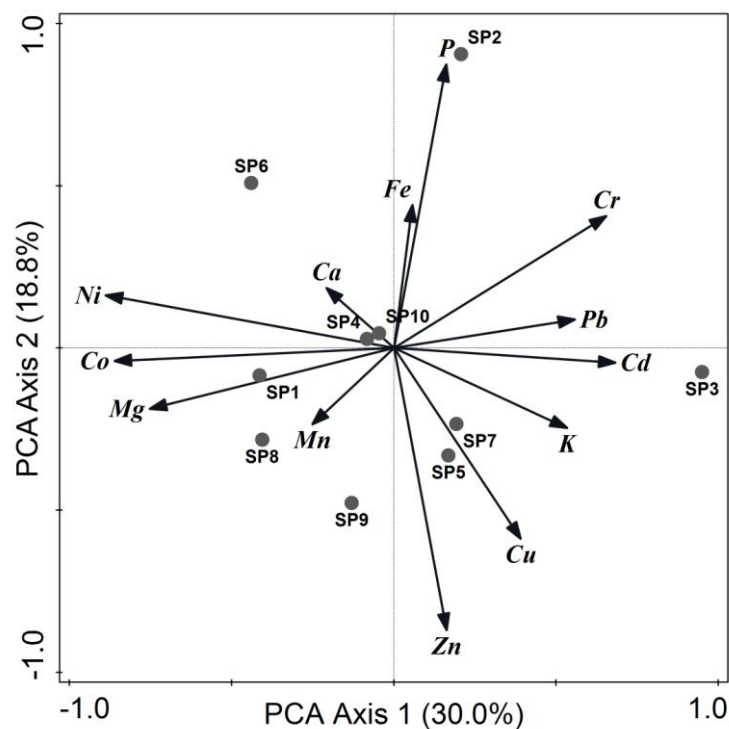


Figure 2. PCA biplot combining data on soil element concentrations from ten serpentine locations in Bosnia and Herzegovina. The first two axes account for 48.8% of the total variance are shown

Table 5. Correlation between elements concentrations in roots and shoots of *Alyssum murale* (the upper right part refers to the shoots and the lower left to the roots). Significant correlation coefficients are in bold (* $p < 0.05$; ** $p < 0.01$ and *** $p < 0.001$)

	Ca	Cd	Co	Fe	K	Mg	Mn	Ni	Pb	Zn
Ca		0.04	0.36*	-0.27	-0.62***	-0.21	0.42*	-0.21	-0.48**	0.71***
Cd	0.36		0.19	-0.13	-0.32	0.47**	0.16	-0.32	-0.43*	-0.33
Co	-0.02	0.27		-0.15	-0.05	0.70***	-0.29	0.24	0.11	0.09
Fe	-0.20	-0.48*	-0.57***		-0.16	0.17	-0.36	-0.21	-0.15	0.25
K	-0.10	-0.07	0.58***	-0.50**		0.01	-0.23	0.57**	0.72***	-0.58***
Mg	-0.32	-0.18	0.10	0.15	-0.12		-0.44*	0.14	0.08	-0.32
Mn	0.16	-0.25	-0.40*	0.24	-0.20	-0.55**		-0.11	-0.15	0.27
Ni	0.14	-0.08	0.58***	-0.06	0.53**	-0.12	-0.07		0.59***	-0.25
Pb	-0.13	-0.12	0.64***	-0.23	0.64***	-0.28	-0.05	0.56**		-0.39*
Zn	-0.16	-0.12	-0.46*	0.59***	-0.57**	-0.07	0.52**	-0.13	-0.48**	

Table 6. Correlation between available elements concentrations in soil and plants tissues (root, stem and leaves) of *Alyssum murale*. Significant correlation coefficients are in bold (* $p < 0.05$; ** $p < 0.01$ and *** $p < 0.001$)

	Ca	Cd	Co	Fe	K	Mg	Mn	Ni	Pb	Zn
roots	0.23	0.13	0.18	0.16	-0.06	-0.38*	0.58***	0.04	0.41*	-0.77***
stems	0.63***	-0.48**	-0.54***	-0.07	0.00	-0.17	0.28	0.14	0.08	-0.66***
leaves	0.64***	-0.14	-0.72***	-0.09	0.22	-0.18	0.60***	0.29	0.22	-0.15

Table 7. Bioconcentration factor (BCF) and translocation factor (TF) of *Alyssum murale*

	Ni	Zn	Mn	Pb	Cd	Co
BCF (shoot/soil)						
SP1	6.48	0.65	0.20	-	10.06	23.19
SP2	9.03	68.39	0.14	-	2.86	6.33
SP3	22.57	4.46	0.25	2.73	-	6.90
SP4	1.93	20.44	0.32	0.00	-	0.81
SP5	5.43	0.84	0.21	-	-	0.50
SP6	6.72	28.08	0.49	-	-	1.98
SP7	7.28	1.48	0.29	49.41	9.67	1.29
SP8	7.02	0.08	0.35	-	-	1.73
SP9	7.96	3.73	0.13	7.60	8.75	1.03
SP10	8.82	12.78	0.20	0.41	3.01	0.67
TF (shoot/root)						
SP1	12.88	0.57	3.03	3.43	2.81	7.05
SP2	5.65	0.57	3.43	2.17	3.38	3.26
SP3	4.22	0.23	4.06	5.05	7.11	11.86
SP4	3.70	0.29	11.39	-	2.75	4.48
SP5	3.71	0.45	5.52	10.66	3.72	1.36
SP6	8.70	0.32	2.33	3.73	5.81	8.11
SP7	3.77	0.59	9.22	5.39	1.60	0.62
SP8	6.33	0.41	7.51	2.61	6.45	2.26
SP9	3.48	2.39	4.81	1.31	1.11	0.98
SP10	3.05	0.25	3.91	1.07	2.86	1.04

Discussion

According to our results ten serpentine soil samples from central Bosnia and Herzegovina have typical ultramafic composition, with the moderate-to-high concentration of available Mg, low-to-moderate available Ca content and elevated total concentrations of trace elements such as Ni, Mn, Co, and Cr. Moreover, the association of elements originating from ultramafic bedrock which occur in high concentrations in these soils is also confirmed by the results of principal component analysis. Regarding threshold values of trace metals which are typical for ultramafic soils Kabata-Pendias et al. (2011) indicated that total Ni concentration of these soils are normally in the range 1400-2000 mg/kg, Co content is from 35 to 200 mg/kg, Cr from 170 to 3400 mg/kg, while total Mn content of serpentine soils differ from 411 to 550 mg/kg.

Results of our study showed the Ni content in the analysed soils of central Bosnia and Herzegovina are very high and within the range of the previously detected values for ultramafic soil samples from other Balkan Peninsula locations, Turkey, and Iran. The Co concentrations at our locations are slightly higher (Shallari et al., 1998; Ghaderian et al., 2007; Reeves and Adigüzel, 2008; Bani et al., 2010; Kazakou et al., 2010; Tumi et al. 2012; Tomović et al., 2013). Taking into account that all the sampling locations are situated far from the households, mainly close to the roads, thus it will be very easy to harvest plants which grow there, and they are excellent candidate sites for Ni phytomining (Tappero et al., 2007; Bani et al., 2015; Van der Ent et al., 2015; Nkrumah et al., 2016). On the other side, total Mn concentration in some other serpentine soils of Serbia was significantly higher, while total Cr value was much higher than on other Balkan peninsula serpentinite locations (Obratov-Petrović et al., 2008; Reeves et al., 2008; Bani et al., 2010) than investigated in our study.

Concerning the trace metal accumulation potential of *A. murale* from Bosnia and Herzegovina, the very high efficiency of Ni uptake is presented. Namely, the content of Ni in analysed tissue samples from Bosnia was variable, where the highest concentration was detected in leaves, and the lowest in the roots. Similar situation was confirmed in Serbian population of *A. murale* where significant interpopulation inconsistency in Ni concentrations among analyzed populations in all plant tissues is shown (Tumi et al., 2012). Generally, in all the plant samples studied concentration of Ni was higher in the shoots compared to the roots. This could be endorsed to easy translocation of Ni through the *A. murale* tissues, which is confirmed by the high values of translocation factor. According to Broadhurst and Chaney (2016), Ni is accumulated mainly in the leaves, especially in vacuoles of epidermal cells and trichome pedicels of *A. murale* population from Kotodesh in Albania. Although, Ni contents in Bosnian *A. murale* leaves (up to 10552.4 mg/kg) are lower than those recorded by Bani et al. (2010) in populations of *A. murale* from Albania (up to 20100 mg/kg), this value is still in the range of those characteristic of Ni hyperaccumulators (Van der Ent et al., 2013). Therefore, the Bosnian population of *A. murale* is one of the best possible candidates for Ni phytomining at the local scale.

The high values of bioconcentration factor in Bosnian population of *A. murale* were also detected for Co. As in the case of Ni, the obtained values of translocation factor for Co were also high, thus the highest concentration of this element was detected in leaves. Although the Co content in leaves of Bosnian population of *A. murale* (up to 142.9 mg/kg) were under a threshold value for Co hyperaccumulators (1000 mg/kg), it was significantly higher than those recorded in populations of *A. murale* from other Balkan countries (Bani et al., 2010; Tumi et al., 2012). This may be explained by the slightly higher Co concentrations of Bosnian soils, given that Tappero et al. (2007) have shown that *A. murale* could accumulate high concentrations of Co in experimental conditions in which nutrient solutions were Co enriched. However, unlike Ni which accumulates mainly in epidermal cells, Co is stored at the leaf tips/margins (Tappero et al. 2007).

Results obtained in this study show that Mn content in soil samples was significantly higher than in plant tissues samples, thus having a low bioconcentration factor, as in the case of *A. murale* from other Balkan countries (Bani et al., 2010; Tumi et al., 2012). The lowest Mn concentration was detected in *A. murale* roots, while the highest Mn content was confirmed in leaves. Zn content has similar trend in plant tissue and soils samples as Mn content. The low Zn content in plant tissue samples, especially in leaves, compared to the Zn content which was significantly higher in soil samples. Tappero et al. (2007) recorded the same trend for Zn in experimental conditions, i.e. moderate values of bioconcentration factor and low values of translocation factor. Moreover, *A. murale* does not accumulate Zn, even when the nutrient solution is enriched with this element (Tappero et al. 2007).

Conclusion

In this study ten soil samples collected from the central Bosnia and Herzegovina showed high level of Ni, Mn, Co and Cr microelements. The relationship between Mg and Ca in all ten soil samples was relatively high as expected and proven in previous studies. Ca concentration, as one of the most important macroelements, was relatively low in soil samples, while in all investigated plant tissue samples of *A. murale* Ca content was significantly higher which also correlates with previous studies. In all

investigated tissue samples the Ca content was higher than Mg content as expected from serpentine soil weathered from serpentinite rock. The concentration of Ni in tissues of all investigated plant samples was high to extremely high while in one case it was 10 times higher in shoot than those in corresponding soil samples derived mainly of ultramafic origin.

According to previous study, metal hyperaccumulators are defined as plants that have the ability to accumulate Ni to such a level that dry leaf concentrations exceeds 1000 mg/kg (0.1 %), so all *A. murale* plant samples from Bosnia and Herzegovina appear as strong Ni hyperaccumulators. Bearing in mind that all *A. murale* plant tissue investigated in our study share the same high capacity for accumulation of Ni and generally shared high tolerance to other metals, these plants could be very good candidates for application in phytoextraction process for phytoremediation and, especially, Ni phytomining.

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